

Conformational conversion of the recombinant hamster prion protein SHaPrP⁹⁰⁻²³² studied by Fourier transform infrared spectroscopy and dynamic light scattering

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The prion protein (PrP) is thought to be responsible for causing transmissible spongiform encephalopathies (TSE), which are neurodegenerative disorders such as BSE in cattle, scrapie in sheep and Creutzfeldt-Jakob disease in humans. The essential step in the transmission and manifestation of these prion diseases seems to be the misfolding of benign cellular PrP (PrP^c) to pathogenic PrP (PrP^{Sc}). Whereas PrP^c has a mainly α -helical structure, PrP^{Sc} seems to be predominantly folded in β -sheets. PrP^{Sc} has the strong tendency to accumulate in neural tissue and to aggregate into insoluble amyloid fibrils.

To study the folding properties of PrP in vitro we expressed a his-tagged fragment of recombinant Syrian hamster PrP^c (amino acids 90–232) in *Escherichia coli* as inclusion bodies. The protein was solubilized in guanidinium hydrochloride (GdnHCl), purified on nickel agarose, and oxidized. After further purification by HPLC it was examined by Fourier Transform Infrared (FT-IR) spectroscopy and dynamic light scattering (DLS) as a function of denaturant (GdnHCl) concentration, ionic strength, and pH.

By analyzing the amide I and amide II region of the FT-IR spectra, our findings show that the secondary structure of PrP is converted from mainly α -helical (bands at 1651 and 1550 cm⁻¹, respectively) to mainly β -sheet (bands at 1619 and 1540 cm⁻¹, respectively) at low pH (4.2) and at either 1–2 M GdnHCl or at sodium chloride concentrations of 0.8–2 M. In contrast, at the same concentrations of GdnHCl or sodium chloride at pH 7.0 such a conversion cannot be observed.

Light scattering measurements show a two phase growth of large PrP aggregates at pH 4.2 and 1 M GdnHCl via a temporarily stable oligomeric intermediate with a 16-fold mass of the monomeric PrP.